

## Synthetic Model of a Bleomycin Metal Complex

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A simple analogue of the metal-complexing pseudopeptide bleomycin has been synthesised and the e.s.r. spectral parameters of its  $\text{Cu}^{\text{II}}$ -complex were found to correspond to the characteristics of the  $\text{Cu}^{\text{II}}$ -bleomycin system.

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Metal-binding properties of the glycopeptide antibiotic anti-tumor agent bleomycin have been the subject of extensive

investigation.<sup>1</sup> The structures of different metallobleomycins have been studied, in particular that of the cupric complex,

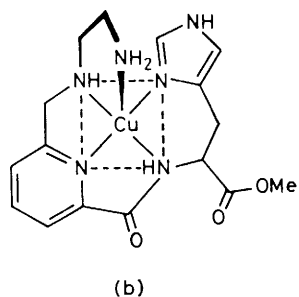
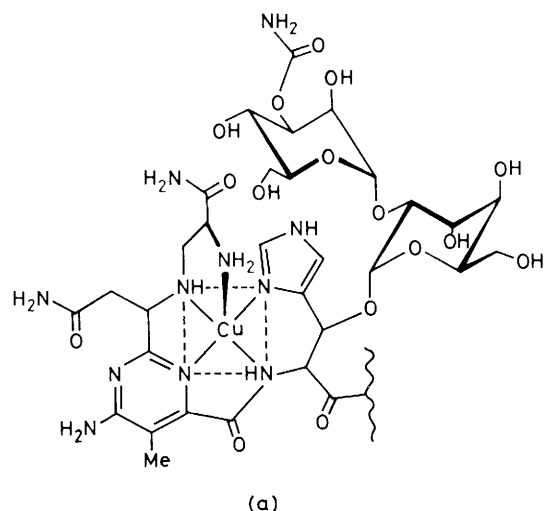
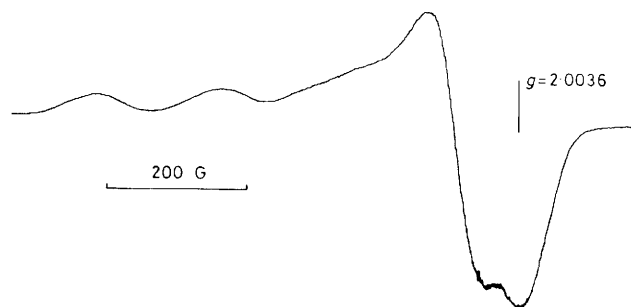
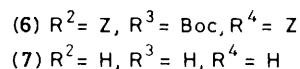
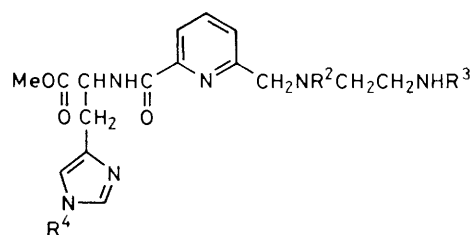
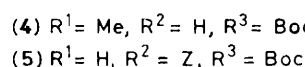
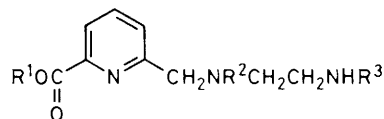
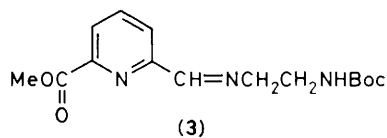
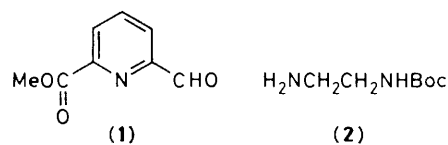


Figure 1

which is the natural form of the drug that is isolated,<sup>2</sup> and the ferrous complex, which has been proposed as the form responsible for the DNA strand scission.<sup>3</sup> On the basis of spectroscopic determinations<sup>4</sup> on transition-metal complexes, in particular on Cu<sup>II</sup>-bleomycin, it has been postulated that the co-ordinating atoms consist of four nitrogen atoms of the pseudopeptide chain in a square plane, with a nitrogen atom of the terminal amine as a fifth axial ligand (Figure 1a). However, alternative structures for Cu<sup>II</sup> or Fe<sup>II</sup>-bleomycin systems have been proposed involving other atoms.<sup>5,6</sup>

We decided to synthesise a simplified model containing the five assumed metal binding sites but excluding all the nitrogen atoms not involved in the chelation.<sup>4</sup> For this purpose, the pyrimidine moiety was replaced with a pyridine ring, substituted by an aminomethyl group instead of by the  $\beta$ -aminopropionamide chain, and the terminal alaninamide group of bleomycin was changed to an aminoethyl side chain. Moreover, the sugar moiety was omitted and  $\beta$ -hydroxy-histidine was simplified to a histidine residue [Figure 1(b)].

Our synthetic strategy for the elaboration of this pseudo-peptide was based on the key compound methyl 6-formylpyridine-2-carboxylate<sup>7</sup> (1) which was combined in ethyl ether at room temperature for 3 h with the appropriate monoprotected ethylenediamine (2) to give the corresponding imine (3). Starting material (2) was prepared in 77% yield by catalytic hydrogenation of (t-butoxycarbonyl)aminoacetonitrile [ $\beta$ -aminoacetonitrile treated with di-*t*-butyl dicarbonate (Boc)<sub>2</sub>O under the classical conditions<sup>8</sup>], in the presence of 5% Raney Ni [b.p. at 0.1 mmHg 72–80 °C,  $\nu_{\max}$  1680 cm<sup>-1</sup>,  $\delta$  (CDCl<sub>3</sub>) 1.32 (2H, s, NH<sub>2</sub>), 1.46 (9H, s, CH<sub>3</sub>), 2.67–3.37 (4H, m, CH<sub>2</sub>), and 5.70 (1H, m, NH)].

Figure 2. E.s.r. spectrum of the model Cu<sup>II</sup>-complex.

Catalytic reductive conversion of the Schiff base (3) [92%,  $\nu_{\max}$  1690 cm<sup>-1</sup>,  $\delta$  (CCl<sub>4</sub>) 1.39 (9H, s, CH<sub>3</sub>), 3.10–3.90 (4H, m, CH<sub>2</sub>), 3.96 (3H, s, CH<sub>3</sub>), 7.67–8.30 (3H, m, pyridyl), and 8.41 (1H, s, CH)] in the presence of Pd-C gave (4) [95%,  $\nu_{\max}$  1705 cm<sup>-1</sup>,  $\delta$  (CDCl<sub>3</sub>) 1.39 (9H, s, CH<sub>3</sub>), 3.10–3.92 (6H, m, CH<sub>2</sub>), 4.00 (3H, s, CH<sub>3</sub>), and 7.57–8.30 (3H, m, pyridyl)] whose secondary amino-function was protected with the benzyloxycarbonyl group Z (by treatment with benzyl chloroformate in the presence of 1 M NaOH in dichloromethane at 0 °C) and whose methyl carboxylate function was hydrolysed (NaOH in methanol–water medium for 2 h at room temperature and neutralization by HCl). The resulting *N,N'*-di-protected free acid (5) (58% from (4),  $\nu_{\max}$  1700 and 1720 cm<sup>-1</sup>,  $\delta$  [(CD<sub>3</sub>)<sub>2</sub>SO] 1.39 (9H, s, CH<sub>3</sub>), 3.15–4.05 (6H, m, CH<sub>2</sub>), 5.14 (2H, s, CH<sub>2</sub>), 7.37 (5H, s, arom.), and 7.74–8.29 (3H, m, pyridyl)) was coupled with *N*(Im)-carbonyl-L-

histidine methyl ester<sup>9</sup> in the presence of dicyclohexylcarbodiimide in dimethylformamide at 0 °C for 3 h and at room temperature for 12 h, affording the protected pseudopeptide (**6**) (70%,  $\nu_{\max}$  1640 and 1720  $\text{cm}^{-1}$ ,  $\delta$  [(CD<sub>3</sub>)<sub>2</sub>SO] 1.40 (9H, s, CH<sub>3</sub>)). The *N*-protecting groups were removed by action of an HBr-acetic acid solution, followed by neutralization with NaHCO<sub>3</sub>, extraction with dichloromethane, and purification on silica-gel. The ester (**7**) [63%,  $\nu_{\max}$  1630 and 1750  $\text{cm}^{-1}$ ] was then complexed by adding CuCl<sub>2</sub> and the e.s.r. spectrum of the Cu<sup>II</sup>-complex was recorded (Figure 2).

The *g*-values ( $g_{\parallel} = 2.21$  and  $g_{\perp} = 2.05$ ) and the hyperfine constant ( $a_{\parallel} = 177$  G) are consistent with values expected for Cu<sup>II</sup> chelated to a square planar array of ligands<sup>10</sup> and with values obtained with the Cu<sup>II</sup>-bleomycin complex.<sup>11</sup> It can be concluded that our simplified synthetic pseudopeptide is a quite satisfactory model for the study of the metal binding sites of bleomycin. They support the previous results of Iitaka<sup>4</sup> and Dabrowiak<sup>11</sup> and refute the structure proposed by Bereman.<sup>6</sup> The model may be a useful tool in the study of the mechanism of bleomycin.

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## References

- 1 H. Umezawa in 'Antibiotics. Mechanism of Action of Antimicrobial and Antitumor Agents,' eds. J. W. Corcoran and F. E. Hahn, Springer-Verlag, New York, 1975, Vol. III, p. 21; N. J. Oppenheimer, J. C. Dabrowiak, F. T. Greenaway, F. S. Santillo, T. Takita, and Y. Sugiura, in 'Bleomycin. Chemical, Biochemical and Biological Aspects,' ed. S. M. Hecht, Springer-Verlag, New York, 1979, p. 124; J. C. Dabrowiak in 'Metal Ions in Biological Systems,' ed. H. Sigel, Marcel Dekker, New York, 1980, Vol. 11, p. 305.
- 2 H. Umezawa, K. Maeda, T. Takeuchi, and Y. Okami, *J. Antibiot.*, 1966, **19**, 200.
- 3 E. A. Sausville, J. Peisach, and S. B. Horwitz, *Biochem. Biophys. Res. Commun.*, 1976, **73**, 814; J. W. Lown and S. K. Sim, *ibid.*, 1977, **77**, 1150.
- 4 Y. Iitaka, H. Nakamura, T. Nakatani, Y. Muraoka, A. Fujii, T. Takita, and H. Umezawa, *J. Antibiot.*, 1978, **31**, 1070; T. Takita, Y. Muraoka, T. Nakatani, A. Fujii, Y. Iitaka, and H. Umezawa, *ibid.*, p. 1073; J. C. Dabrowiak, F. T. Greenaway, and R. Grulich, *Biochemistry*, 1978, **17**, 4090; C. M. Vos, G. Westra, and D. J. Schipper, *J. Inorg. Biochem.*, 1980, **13**, 165; M. Otsuka, M. Yoshida, S. Kobayashi, M. Ohno, Y. Sugiura, T. Takita, and H. Umezawa, *J. Am. Chem. Soc.*, 1981, **103**, 6986.
- 5 N. J. Oppenheimer, L. O. Rodriguez, and S. M. Hecht, *Proc. Natl. Acad. Sci. U.S.A.*, 1979, **76**, 5616.
- 6 R. D. Bereman and M. E. Winkler, *J. Inorg. Biochem.*, 1980, **13**, 95.
- 7 W. Mathes, W. Sauermilch, and T. Klein, *Chem. Ber.*, 1953, **86**, 584.
- 8 L. Moroder, A. Hallett, E. Wunsch, O. Keller, and G. Wersin, *Hoppe-Seyler's Z. Physiol. Chem.*, 1976, **357**, 1651.
- 9 K. Inouye and H. Otsuka, *J. Org. Chem.*, 1962, **27**, 4243.
- 10 J. Peisach and W. E. Blumberg, *Arch. Biochem. Biophys.*, 1974, **165**, 691.
- 11 J. C. Dabrowiak, F. T. Greenaway, W. F. Longo, M. Van Husen, and S. T. Crooke, *Biochim. Biophys. Acta*, 1978, **517**, 517.